

Primary Pulmonary Ewing Sarcoma or Askin Tumor (PNET of the Thoracopulmonary Region) - a Rare Clinical Case in our Practice

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Abstract

Primary pulmonary Ewing sarcoma is a rare disease belonging to the Ewing sarcoma tumor family. The origin of these rarely diagnosed primary undifferentiated neoplasms with possible localization in different organs and tissue structures is still unknown. We present a rare clinical case of primary pulmonary extraskeletal Ewing sarcoma in a woman 35 years old. The purpose of this article is to emphasize the difficult histological and differential diagnosis, requiring a wide panel of immunohistochemical markers. Despite the maximum possible radical surgery, given the local biological malignancy of this exceptionally rare tumor with a poor prognosis, we performed postoperative intensity modulated radiotherapy (IMRT).

Keywords: Primary pulmonary ewing sarcoma, Extrasosseous primitive neuroectodermal tumor (PNET), Immunohistochemistry, PET/CT, Surgery, Radiotherapy

1. Introduction

Ewing's sarcoma family tumors (ESFT) includes Ewing sarcoma (osseous and extrasosseous), primitive neuroectodermal tumor (PNET) of the bone and soft tissues, Askin tumor (PNET of the thoracopulmonary region) [1]. Extraskeletal Ewing's sarcoma (EES) has also been reported in the pelvis, buttocks, upper limbs, mediastinum, and abdominal organs [2]. Pulmonary EES is rare, and fewer than 30 pulmonary PNET cases have been reported thus far [1,3]. Immunohistochemistry is compulsory to confirm the diagnosis [4]. We present a rare clinical case of primary pulmonary EES in a woman 35 years old. The purpose of this article is to emphasize the difficult diagnosis of this rare disease at a young age and the individual assessment of the necessary therapeutic approach.

2. Clinical Case

We present a 35-year-old woman. On the occasion of a complaint of subjective shortness of breath in April 2024, a lung CT was carried out on which a rounded lesion was found with a sharp outline in the fifth segment of a left lung measuring 52 x42 mm suspected for an echinococcal cyst. In September 2024 she entered the thoracic surgery clinic, where a video-assisted partial resection of the left lung was performed. Intraoperatively- Through two thoracoports to the left reached the left pleural cavity. The lung is free from adhesions. In the lower part, a cystic pulmonary

formation is located, which is partially resisted. A biopsy of the thoracic pleura was performed.

Histological Result: Pulmonary parenchyma fragments with infiltration of malignant tumor tissue, made up of monotonous moderately large cells with a moderate amount of cytoplasm and large amorphous nucleus with stranded eosinophilic nucleol. The cells are arranged in solid sheets, in places perivascular, similar to neuroectodermal differentiation (Figure 1). The tumor complies with the underlying pulmonary parenchyma.

Immunohistochemistry: Neoplastic cells expose diffusely CD99 (Figure 2 A), Vimentin (Figure 2 B), CD56 (Figure 2 C), ERG and Beta -catenin membrane. Single tumor cells expose Synaptophysin (Figure 2 D). Negative expression for MCK, CK7 (Figure 3 A), CK20 (Figure 3 B), EMA, TTF-1 (Figure 3 C), HMB -45 (Figure 3 D), S100, AR, WT-1, POSE, PAX-5, SMA and P40 CD20, CD25, CD2, CD4, CD5, CD3C (positive in peritumoral plasmocytes), CD31 and CD34 (positive focal in endothelium). The proliferative activity measured by KI 67 is about 25-30% (Figure 4).

Conclusion: Morphology and immunofenotype of neoplasm corresponds to primary pulmonary Ewing sarcoma with a specific genotype. From the preoperative PET/CT-neoplasm in the fifth segment of the left lung, which touches the bone pleura. Other

metabolite active lesions with the type of malignant engagement are not scanned.

Primary pulmonary Ewing sarcoma without locoregional and distant dissemination (Figure 5). In November 2024, a lateral thoracotomy was performed on the left with a massive radical resection of ewing sarcoma in a block with the adjacent part of the fifth rib and atypical resection of the lingula of the upper lung lobe. Intraoperatively, a 7x 8 cm neoplasm is found /with preliminary histological data for Ewing sarcoma /, intimately adjacent to the fifth rib, infiltrating the upper lung lobe in the lingula area. The neoplasm described was removed as radically as possible with the adjacent part of the fifth rib, as well as atypically with the adjacent part of the lingula. The Hilus lymph node is extirpated.

Histological Diagnosis-Macroscopically-Two Containers with

Inscription: 1. "Lung"- a 9 cm long rib and associated with it a pulmonary parenchyma of almost spherical shape and a diameter of 7 cm. In the cut almost completely engaged by a pink red tumor with a homogeneous measurement surface. 2. "Hilar lymph node" with a diameter of 12x7 mm.

Morphological result: A tumor composed of cells with unclear cytoplasmic borders, the nuclei are of a fine priest chromatin, the cytoplasm is pale eosinophilic. The cells form layers and wide cords. The stroma is scarce and richly vascularized by thin-walled blood vessels (Figure 6). The tumor is well distorted by the adjacent pulmonary parenchyma of a fibrous capsule. Part of the rib with a small fragment of the tumor in the bone marrow, but

without a direct tumor infiltration. The hilus lymph node is without metastasis with sinus histiocytosis and anthracosis (Figure 7).

Conclusion: Lung engaged by probable Ewing sarcoma. Immunohistochemical (IHC) verification is required. CK AE1/3 with a negative reaction in tumor cells with positive internal control in the pulmonary parenchyma; Synaptophysin- a positive reaction in scattered single tumor cells (Figure 8 A); FLI-1- weakly to moderately pronounced reaction in about 70% of tumor cells; CD99- diffuse positive reaction in almost 100% of tumor cells (Figure 8 B); CD 56- Positive reaction in about 85-90% of tumor cells (Figure 8C); CD 45 with a negative reaction to tumor cells (Figure 8 D) with positive internal control in lymphocytes; SOX 10 (Figure 8 E), Chromogranin A (Figure 8 F), TLE and TDT- with a negative reaction. *Postoperative PET/CT-* Without data on residual tumor tissue, as well as data on lymphogenic and hematogenous dissemination (Figure 9). To reject the extraskeletal Ewing sarcoma of the uterus, the patient did ultrasound and MRI of the uterine body with the following finding: a few small uterine myomas (Figure 10). Due to the nearby resection lines, the local aggressive expansion of the extraskeletal Ewing sarcoma and the young age of the woman, we estimated that it was necessary to conduct a postoperative additive radiotherapy (RT). Figure 11 shows the targeted volumes in the overlapping (fusion) of the images from the postoperative PET/CT of the lungs with that of the planning CT. We conducted an intensity modulated radiotherapy of the VMAT method with daily dose (DD) 1.8 Gy up to total dose (TD) 45 Gy (Figure 12), which the patient suffered without acute side effects from the lungs and heart.

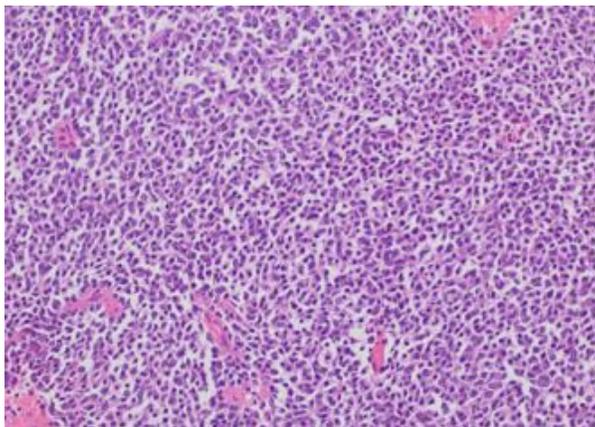


Figure 1: Pulmonary parenchyma fragments with infiltration of malignant tumor tissue, made up of monotonous moderately large cells with a moderate amount of cytoplasm and large amorphous nucleus with stranded eosinophilic nucleol. The cells are arranged in solid sheets, in places perivascular, similar to neuroectodermal differentiation (Hematoxylin-Eosin, original magnification x200)

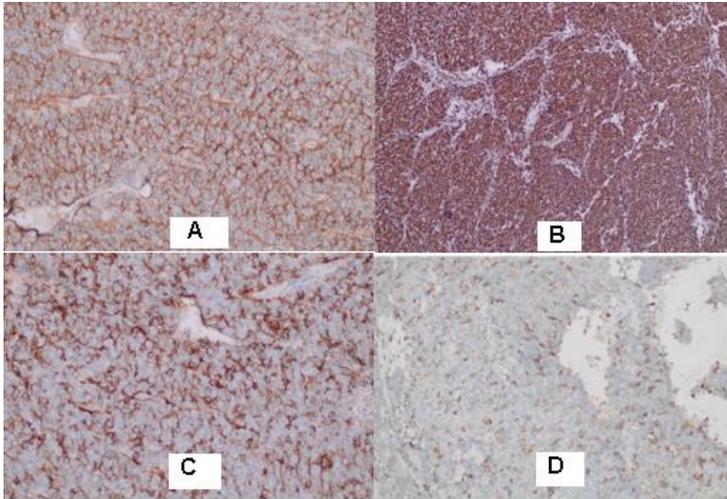


Figure 2: Immunohistochemistry- A) Neoplastic cells with diffuse positive membrane reaction for CD99; B) Positive reaction for Vimentin; C) Positive membrane reaction for CD56; D) Single tumor cells with a positive reaction for Synaptophysin x 100.

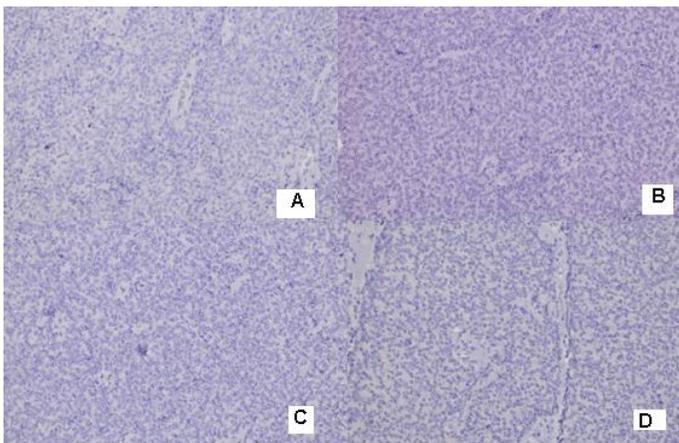


Figure 3: Immunohistochemistry- Negative expression reaction for A) CK7; B) CK20; C) TTF-1; D) HMB -45 x100

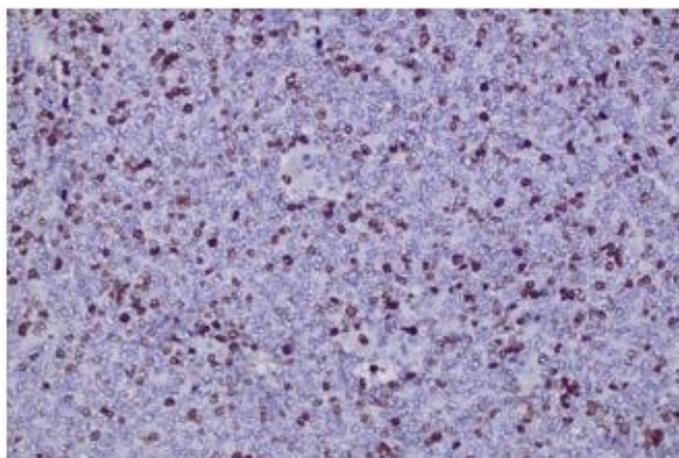


Figure 4: Immunohistochemistry - High level of the proliferation marker Ki 67 25-30% x100

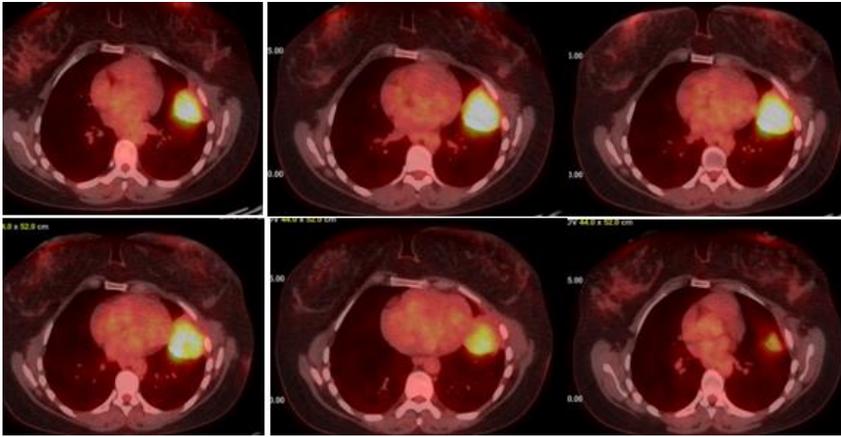


Figure 5: Preoperative PET/CT- Neoplasm in the fifth segment of the left lung, which touches the bone pleura. Other metabolite active lesions with the type of malignant engagement are not scanned

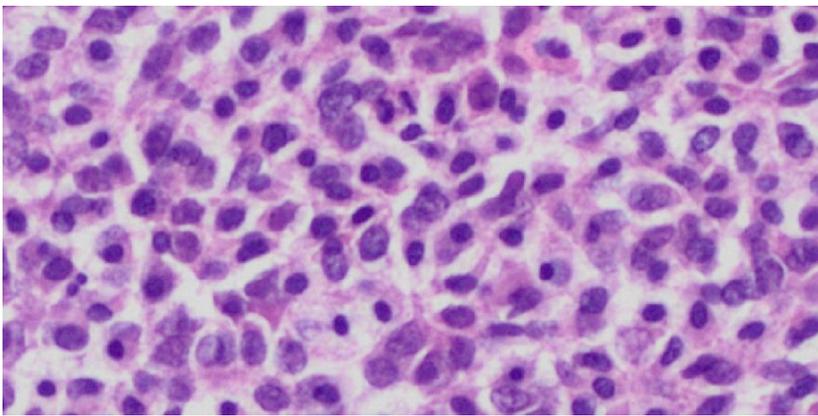


Figure 6: A neoplasm composed of cells with unclear cytoplasmic borders, the nuclei are of a fine priest chromatin, the cytoplasm is pale eosinophilic. The cells form layers and wide cords. The stroma is scarce and richly vascularized by thin -walled blood vessels. (Hematoxylin-Eosin, original magnification x400)

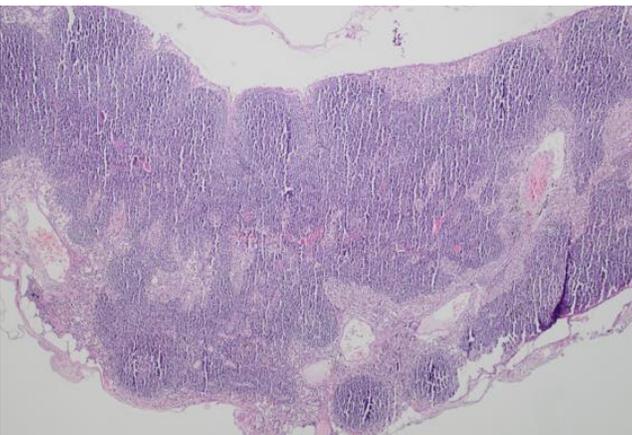


Figure 7: The hilus lymph node is without metastasis with sinus histiocytosis and anthracosis (Hematoxylin-Eosin, original magnification x200)

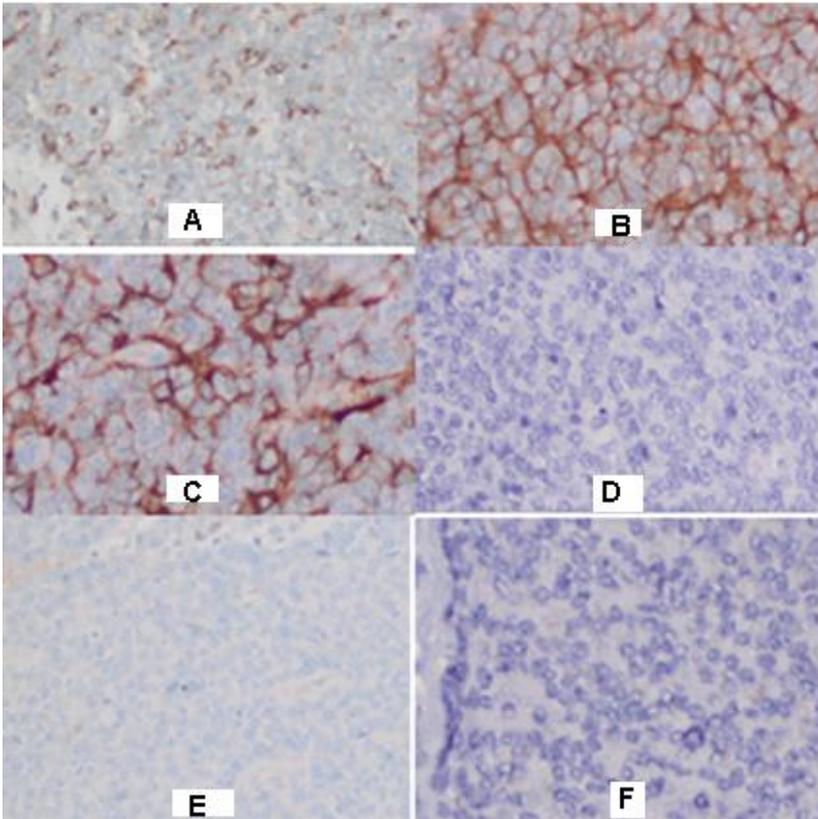


Figure 8: A) Synaptophysin with a positive reaction in scattered single tumor cells x 400; B) CD99 with diffuse positive reaction in almost 100% of tumor cells x 400; C) CD 56- Positive reaction in about 85-90% of tumor cells x 400; D) CD 45 with a negative reaction to tumor cells with positive internal control in lymphocytes x 400; E) SOX 10 with a negative reaction to tumor cells x 400; F) Chromogranin A with a negative reaction to tumor cells x 400

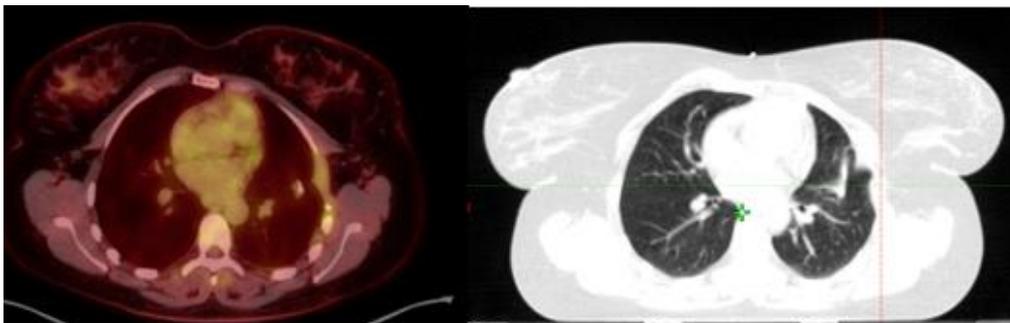


Figure 9: Postoperative PET/CT and CT- Without data on residual tumor tissue, as well as data on lymphogenic and hematogenous dissemination

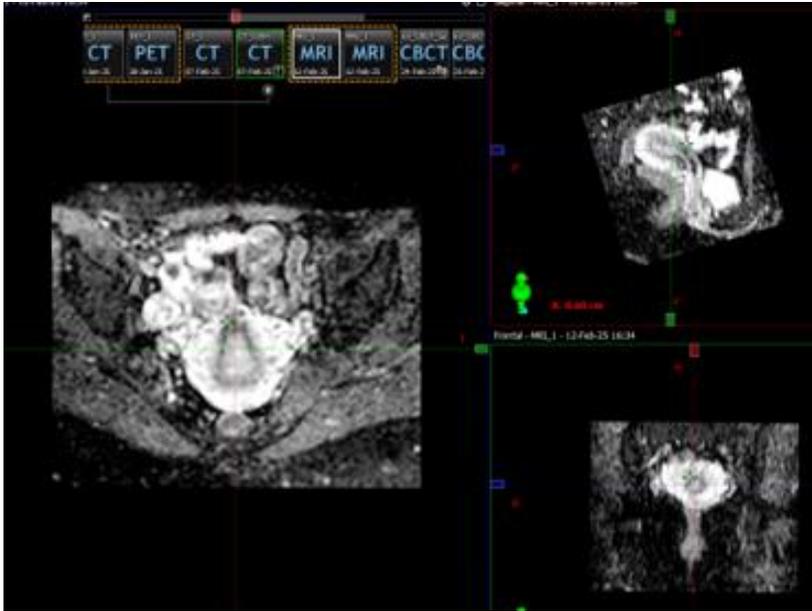


Figure 10: MRI of the uterine body with: a few small uterine myomas.

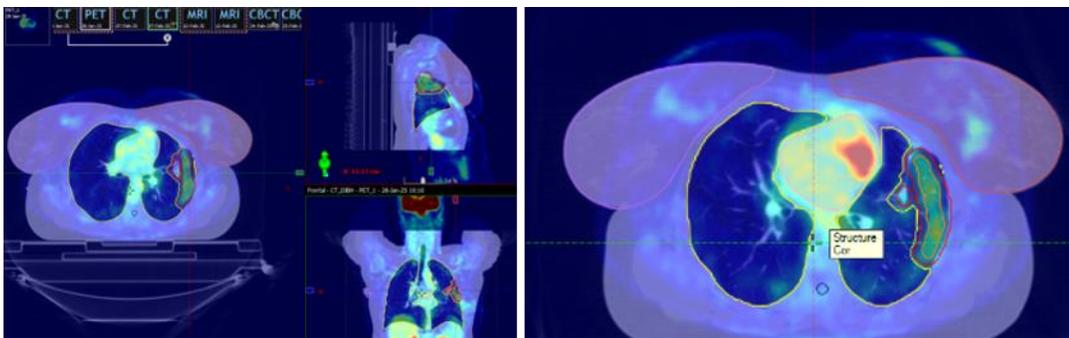


Figure 11: The targeted volumes in the overlapping (fusion) of the images from the postoperative PET/CT of the lungs with that of the planning CT

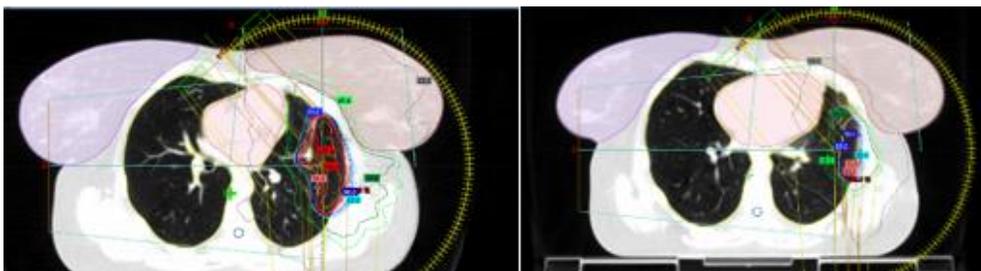


Figure 12: Intensity modulated radiotherapy (IMRT) of the VMAT method In the area of suboptimal resection lines with daily dose (DD) 1.8 Gy up to total dose (TD) 45 Gy. To reduce the dose in the heart, radiation is performed with deep respiratory inspiration.

3. Discussion

Extraskelatal Ewing sarcoma (EES) is a rare entity that belongs to the ES family of tumors (ESFT), which is a group of small round tumor cells that share a common neural histology and genetic mechanism [5-8]. The ESFT encompass all the previous terminologies such as Ewing sarcoma of bone, extra-skeletal Ewing sarcoma, peripheral primitive neuroectodermal tumour,

Askin tumour of thoraco-pulmonary region, and atypical Ewing sarcoma based on the presence of a unifying pathognomonic chromosomal translocation [9-18].

The ESFT are an entity composite of malignant small round blue cell tumors with different degrees of neuroectodermal differentiation [19,20]. They are characterized by strong continuous cell membrane

expression of CD99 protein, and genetic translocation involving Ewing's sarcoma (EWS) in 90% of cases [21]. Pathological findings typically reveal the presence of small round cells characterized by poor differentiation, hyperchromatic nuclei, high nuclear-cytoplasmic ratios, and the formation of Homer-Wright rosettes [22]. By 2020, 34 patients with ESFT were published in English literature in various organs (ovary, cervix, vagina, kidneys, bladder, breast, brain) of which 4 patients with primary pulmonary ES. The median age of patients with lung involvement was 33.5 (range: 32-41)years. Immunohistochemistry for CD99 and FLI1 was positive in all 4 [23]. The disease arises from some mesenchymal or neural crest-derived stem or progenitor cell and hence may show variable neuronal, mesenchymal, and epithelial differentiation [10,14,24]. The diagnosis of EES is mainly based on histopathology and immunohistochemistry of tumor specimens. Under a light microscope, the tumor cells are small and round, uniform in size with a round to oval in the nucleus, and sparse in the cytoplasm. Tumor cells are arranged in flakes or lobules, which are separated by vascular fibrous tissues [25]. Figure 1 and Figure 6 shows monotonous moderately large cells with unclear cytoplasmic borders, the nuclei are of a fine priest chromatin, the cytoplasm is pale eosinophilic. The cells form layers and wide cords. The stroma is scarce and richly vascularized by thin-walled blood vessels. Neoplastic cells do not form the rosettes of Homer-Wright. EES shows cytologic features similar to those of its skeletal counterpart, with a continuum from the typical/small cell form in the majority of cases to the intermediate and atypical/large cell variants in a minority of cases [26]. The large cell pattern, also known as "atypical Ewing sarcoma," features large cells with prominent nucleoli, a mild degree of nuclear pleomorphism, and eosinophilic cytoplasm, which may lead to rhabdoid cells [27]. Immunohistochemical (IHC) staining demonstrated positive membranous expression of CD99, CD56, Vimentin, Synaptophysin and high levels of the Ki67 proliferation marker [22]. Membranous CD99 and nuclear Fli-1 staining was seen in 10 and 16 tumors, respectively, and concurrent expression of both markers was seen in both central and Ewing sarcoma/peripheral PNETs [28]. Such a typical immunohistochemistry is also reported in our clinical case in Figure 2, Figure 4 and Figure 8/ ABC, but only single cells take into account a positive immunohistochemical reaction to Synaptophysine (Figure 2/D and Figure 8/A), which is indicative of neuroectodermal differentiation. By immunohistochemistry, 17 PNETs expressed at least 1 marker of neuronal differentiation, including synaptophysin, NSE, CD56, S100, and chromogranin in 10, 8, 14, 8, and 1 tumor, respectively [28]. Ewing sarcomas are often positive for one or more neuroendocrine immunohistochemical stains, such as synaptophysin and neuron-specific enolase, which highlights areas of neuroectodermal differentiation present in these tumors [29,30]. Antibodies against CD56 may be useful in the identification of neuroendocrine malignancies, especially if the cells do not express CD45, indicating a non-leucocyte origin [31]. A similar immunohistochemical reaction to the CD56 and CD 45 is found in the clinical case presented (Figure 8/C and Figure 8/D). Histological morphology of tumor cells together with the immunohistochemical analysis support the final diagnosis of

cytologically atypical primary pulmonary Ewing sarcoma with moderate neuroectodermal differentiation.

CT showed the location, size, and internal tissue structure of the tumor, which is of great importance in evaluating the feasibility of tumor resection and formulating the treatment plan [25]. Magnetic resonance (MR) imaging is performed prior to biopsy to help determine the optimal biopsy site and to avoid the distortion caused by post biopsy changes [32]. MRI and fluorodeoxyglucose positron emission tomography (FDG PET) imaging are used for initial diagnosis and detection of metastasis, respectively [33-35]. Gynecologic PNETs have been reported in the ovary [36], broad ligament [37], uterine corpus [38], uterine cervix [39], vagina [40], and vulva [41]. In a differential diagnostic plan on the presence of a PNET with a gynecological localization and the presence of a solitary pulmonary metastasis, we decided that uterine MRI was needed (Figure 10). 18-Fluorodeoxyglucose positron emission tomography showed that Ewing sarcoma of the lung often showed large volume, smooth edge, and high fluorodeoxyglucose uptake [42]. In Figure 3, we present the preoperative PET/CT in our patient, which shows a large rounded formation in the left lung with high metabolic activity, touching to the 5th rib. Intraoperatively turns out that the tumor is surrounded by a fibrous capsule that distinguishes it from the surrounding pulmonary parenchyma, and from the pathohistological result, the resection lines prove to be extirpated in a healthy. The presence of a fibrous capsule around the tumor, as well as the large amount of plasmocytes, B cells, and T cells around the tumor cells, determines the high level of autoimmune reaction in the presented clinical case (Figure 6). Previously, EES was treated using the soft tissue sarcomas protocol [43]; however, the current treatment recommendation by the National Comprehensive Cancer Network (NCCN) is local treatment (surgery and/or radiotherapy) plus chemotherapy [44,45].

Management of primary pulmonary EES/PNET usually needs aggressive multimodality treatment with upfront generous surgery followed by chemotherapy plus or minus radiotherapy [46]. Considering that EES is a highly invasive malignant tumor, the most effective treatment is surgical resection combined with postoperative chemotherapy and high-dose radiotherapy (RT) [47]. The overall 5 year survival rate is better in patients who undergo complete resection, with wide surgical margins compared with suboptimal margins [48]. Surgery is generally the recommended treatment option for patients with resectable pulmonary PNET, where marginal or wide resection is possible [45,49]. In cases where surgery is not a feasible option, a combination of chemotherapy and radiotherapy should be considered as an alternative treatment approach [22]. The efficacy of adjuvant chemotherapy remains controversial, as it does not result in a significant reduction in the risk of death [50,51]. In situations where complete tumor removal is not achievable, a combination of radiotherapy (RT) or chemoradiation (Ch-RT) is recommended [22]. Figure 9 shows the postoperative PET/CT, which does not take into account a residual tumor, but due to the suboptimal resection margins, as well as

the high proliferative index KI 67 30% (Figure 4), taking into account the malignant nature of this undifferentiated neoplasm, we considered that postoperative RT was required. Another reason for the need for postoperative RT is the initial surgery at which a partial tumor resection was performed, as well as the pathohistological result from the second surgery with a small tumor fragment in the bone marrow in part of the researched rib. We conducted an IMRT of the VMAT method in the area of suboptimal resection lines with daily dose (DD) 1.8 Gy up to total dose (TD) 45 Gy (Figure 12), which the patient suffered without acute side effects from the lungs and heart. Total dose 45 Gy is sufficient for the preventive irradiation of nearby resection linings. The patient was evaluated for active monitoring, namely through a pulmonary MRI after 2 months and PET/ CT after 3 months.

4. Conclusion

The primary lung Ewing sarcoma is an extremely rare neoplasm with published about 35 clinical cases in the world. It is mostly diagnosed at a young age. Due to the large histopathological variation of non -differentiated tumors from the Ewing sarcoma family, a very wide range of immunohistochemical and genetic analysis is required. Due to the extremely aggressive nature of these extraosseous tumors, the main treatment for localized disease without distant metastases is surgical, at discretion accompanied by postoperative RT and/ or chemotherapy. Active monitoring through a pulmonary MRI and PET/ CT is required.

References

1. Asker, S., Sayir, F., Bulut, G., Sunnetcioglu, A., Ekin, S., & Yavuz, A. (2015). Primitive neuroectodermal tumor/ewing sarcoma presenting with pulmonary nodular lesions. *Case reports in oncological medicine*, 2015, 957239.
2. Colovic, R. B., Grubor, N. M., Micev, M. T., Matic, S. V., Atkinson, H. D., & Latincic, S. M. (2009). Perigastric extraskeletal Ewing's sarcoma: a case report. *World journal of gastroenterology*, 15(2), 245–247.
3. Wang, N., Dong, S. S., Wu, C. L., Wang, Y., Meng, L., Ren, Y., Cui, X. B., Li, M., & Qi, Y. (2021). Rare Primary Pulmonary Primitive Neuroectodermal Tumor: A Case Report and Literature Review. *OncoTargets and therapy*, 14, 139–144.
4. Sobh, E., El-Sheshtawy, W.H. & Anis, S.E. Primary pulmonary extraskeletal Ewing sarcoma/Primitive neuroectodermal tumor: Two case reports. *Egypt J Bronchol* 11, 161–164 (2017).
5. van den Berg, H., Heinen, R. C., van der Pal, H. J., & Merks, J. H. (2009). Extra-osseous Ewing sarcoma. *Pediatric hematology and oncology*, 26(4), 175–185.
6. Peabody TD and Attar S (eds): SpringerLink: Orthopaedic Oncology: Primary and metastatic tumors of the skeletal system. In: *Cancer Treatment and Research*. (2014) Cham, Springer International Publishing, Imprint, Springer, Berlin, Gernay, pp 203 223.
7. El Weshi, A., Allam, A., Ajarim, D., Al Dayel, F., Pant, R., Bazarbashi, S., & Memon, M. (2010). Extraskeletal Ewing's sarcoma family of tumours in adults: analysis of 57 patients from a single institution. *Clinical oncology (Royal College of Radiologists (Great Britain))*, 22(5), 374–381.
8. Carpentieri, D. F., Qualman, S. J., Bowen, J., Krausz, T., Marchevsky, A., Dickman, P. S., & Cancer Committee, College of American Pathologists (2005). Protocol for the examination of specimens from pediatric and adult patients with osseous and extraosseous ewing sarcoma family of tumors, including peripheral primitive neuroectodermal tumor and ewing sarcoma. *Archives of pathology & laboratory medicine*, 129(7), 866–873.
9. Delattre, O., Zucman, J., Melot, T., Garau, X. S., Zucker, J. M., Lenoir, G. M., Ambros, P. F., Sheer, D., Turc-Carel, C., & Triche, T. J. (1994). The Ewing family of tumors--a subgroup of small-round-cell tumors defined by specific chimeric transcripts. *The New England journal of medicine*, 331(5), 294–299.
10. Folpe, A. L., Hill, C. E., Parham, D. M., O'Shea, P. A., & Weiss, S. W. (2000). Immunohistochemical detection of FLI-1 protein expression: a study of 132 round cell tumors with emphasis on CD99-positive mimics of Ewing's sarcoma/primitive neuroectodermal tumor. *The American journal of surgical pathology*, 24(12), 1657–1662.
11. O'Sullivan, M. J., Perlman, E. J., Furman, J., Humphrey, P. A., Dehner, L. P., & Pfeifer, J. D. (2001). Visceral primitive peripheral neuroectodermal tumors: a clinicopathologic and molecular study. *Human pathology*, 32(10), 1109–1115.
12. Khoury J. D. (2005). Ewing sarcoma family of tumors. *Advances in anatomic pathology*, 12(4), 212–220.
13. Gamberi, G., Cocchi, S., Benini, S., Magagnoli, G., Morandi, L., Kreshak, J., Gambarotti, M., Picci, P., Zanella, L., & Alberghini, M. (2011). Molecular diagnosis in Ewing family tumors: the Rizzoli experience--222 consecutive cases in four years. *The Journal of molecular diagnostics : JMD*, 13(3), 313–324.
14. Pinto, A., Dickman, P., & Parham, D. (2011). Pathobiologic markers of the ewing sarcoma family of tumors: state of the art and prediction of behaviour. *Sarcoma*, 2011, 856190.
15. de Alava, E., Lessnick, S.L., Sorensen, P.H. (2013). Ewing sarcoma. In: Fletcher CDM, Bridge JA, Hogendoorn PCW, Mertens F, eds. *WHO classification of tumours of soft tissue and bone*. Lyon, France: WHO Press:305-309.
16. Sand, L. G., Szuhai, K., & Hogendoorn, P. C. (2015). Sequencing Overview of Ewing Sarcoma: A Journey across Genomic, Epigenomic and Transcriptomic Landscapes. *International journal of molecular sciences*, 16(7), 16176–16215.
17. Wei, S., Henderson-Jackson, E., Qian, X., & Bui, M. M. (2017). Soft Tissue Tumor Immunohistochemistry Update: Illustrative Examples of Diagnostic Pearls to Avoid Pitfalls. *Archives of pathology & laboratory medicine*, 141(8), 1072–1091.
18. Javalgi, A. P., Karigoudar, M. H., & Palur, K. (2016). Blue Cell Tumour at Unusual Site: Retroperitoneal Ewings Sarcoma. *Journal of clinical and diagnostic research : JCDR*, 10(4), ED19–ED20.
19. Javalgi, A. P., Karigoudar, M. H., & Palur, K. (2016). Blue Cell Tumour at Unusual Site: Retroperitoneal Ewings Sarcoma.

- Journal of clinical and diagnostic research : JCDR, 10(4), ED19–ED20.
20. Lee, Y. Y., Kim, D. H., Lee, J. H., Choi, J. S., In, K. H., Oh, Y. W., Cho, K. H., & Roh, Y. K. (2007). Primary pulmonary Ewing's sarcoma/primitive neuroectodermal tumor in a 67-year-old man. *Journal of Korean medical science*, 22 Suppl(Suppl), S159–S163.
 21. Renard, C., & Ranchère-Vince, D. (2015). Tumeurs de la famille Ewing/PNET : vers un nouveau paradigme ? [Ewing/PNET sarcoma family of tumors: towards a new paradigm?]. *Annales de pathologie*, 35(1), 86–97.
 22. Zhang, Y., Shang, K., Li, J., Sun, M., & Gu, X. (2024). Operative treatment of pulmonary primitive neuroectodermal tumor: a case report and literature review. *Journal of cardiothoracic surgery*, 19(1), 109.
 23. Murthy, S. S., Challa, S., Raju, K., Rajappa, S. J., Fonseca, D., Gundimeda, S. D., Rao, B. V., Ahmed, F., Kodandapani, S., Nambaru, L., Mundada, M. C., Sharma, R., Mallavarapu, K. M., Koppula, V. C., & Rao, T. S. (2020). Ewing Sarcoma With Emphasis on Extra-skeletal Ewing Sarcoma: A Decade's Experience From a Single Centre in India. *Clinical pathology (Thousand Oaks, Ventura County, Calif.)*, 13, 2632010X20970210.
 24. Kovar, H., Amatruda, J., Brunet, E., Burdach, S., Cidre-Aranaz, F., de Alava, E., Dirksen, U., van der Ent, W., Grohar, P., Grünewald, T. G., Helman, L., Houghton, P., Iljin, K., Korsching, E., Ladanyi, M., Lawlor, E., Lessnick, S., Ludwig, J., Meltzer, P., Metzler, M., ... Delattre, O. (2016). The second European interdisciplinary Ewing sarcoma research summit—A joint effort to deconstructing the multiple layers of a complex disease. *Oncotarget*, 7(8), 8613–8624.
 25. Zou, X., Chang, W., & Gao, H. (2021). A primary Ewing's sarcoma of pleura: Case report and literature review. *Respiratory medicine case reports*, 34, 101516.
 26. Guiter, G. E., Gamboni, M. M., & Zakowski, M. F. (1999). The cytology of extraskeletal Ewing sarcoma. *Cancer*, 87(3), 141–148.
 27. Yoshida A. (2023). Ewing and Ewing-like sarcomas: A morphological guide through genetically-defined entities. *Pathology international*, 73(1), 12–26.
 28. Chiang, S., Snuderl, M., Kojiro-Sanada, S., Quer Pi-Sunyer, A., Daya, D., Hayashi, T., Bosincu, L., Ogawa, F., Rosenberg, A. E., Horn, L. C., Wang, L., Iafrate, A. J., & Oliva, E. (2017). Primitive Neuroectodermal Tumors of the Female Genital Tract: A Morphologic, Immunohistochemical, and Molecular Study of 19 Cases. *The American journal of surgical pathology*, 41(6), 761–772.
 29. Ozobokeme OE, Jones TE, Naous R, Khader SN. (2022) Educational Case: Ewing sarcoma family of tumors: Clinical presentation, pathologic findings, and differential diagnosis. *Acad Pathol*. 13;9(1):100051.
 30. Weissferdt, A., Kalhor, N., & Moran, C. A. (2015). Ewing sarcoma with extensive neural differentiation: a clinicopathologic, immunohistochemical, and molecular analysis of three cases. *American journal of clinical pathology*, 143(5), 659–664.
 31. Bryson, G. J., Lear, D., Williamson, R., & Wong, R. C. (2002). Detection of the CD56+/CD45- immunophenotype by flow cytometry in neuroendocrine malignancies. *Journal of clinical pathology*, 55(7), 535–537.
 32. Brisse, H., Ollivier, L., Edeline, V., Pacquement, H., Michon, J., Glorion, C., & Neunschwander, S. (2004). Imaging of malignant tumours of the long bones in children: monitoring response to neoadjuvant chemotherapy and preoperative assessment. *Pediatric radiology*, 34(8), 595–605.
 33. Meyer, J. S., Nadel, H. R., Marina, N., Womer, R. B., Brown, K. L., Eary, J. F., Gorlick, R., Grier, H. E., Randall, R. L., Lawlor, E. R., Lessnick, S. L., Schomberg, P. J., & Kailo, M. D. (2008). Imaging guidelines for children with Ewing sarcoma and osteosarcoma: a report from the Children's Oncology Group Bone Tumor Committee. *Pediatric blood & cancer*, 51(2), 163–170.
 34. Franzius, C., Sciuk, J., Daldrup-Link, H. E., Jürgens, H., & Schober, O. (2000). FDG-PET for detection of osseous metastases from malignant primary bone tumours: comparison with bone scintigraphy. *European journal of nuclear medicine*, 27(9), 1305–1311.
 35. Völker, T., Denecke, T., Steffen, I., Misch, D., Schönberger, S., Plotkin, M., Ruf, J., Furth, C., Stöver, B., Hautzel, H., Henze, G., & Amthauer, H. (2007). Positron emission tomography for staging of pediatric sarcoma patients: results of a prospective multicenter trial. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*, 25(34), 5435–5441.
 36. Ostwal, V., Rekhi, B., Noronha, V., Basak, R., Desai, S. B., Maheshwari, A., & Prabhash, K. (2012). Primitive neuroectodermal tumor of ovary in a young lady, confirmed with molecular and cytogenetic results—a rare case report with a diagnostic and therapeutic challenge. *Pathology oncology research : POR*, 18(4), 1101–1106.
 37. Lee, K. M., & Wah, H. K. (2005). Primary Ewing's sarcoma family of tumors arising from the broad ligament. *International journal of gynecological pathology : official journal of the International Society of Gynecological Pathologists*, 24(4), 377–381.
 38. Euscher, E. D., Deavers, M. T., Lopez-Terrada, D., Lazar, A. J., Silva, E. G., & Malpica, A. (2008). Uterine tumors with neuroectodermal differentiation: a series of 17 cases and review of the literature. *The American journal of surgical pathology*, 32(2), 219–228.
 39. Malpica, A., & Moran, C. A. (2002). Primitive neuroectodermal tumor of the cervix: a clinicopathologic and immunohistochemical study of two cases. *Annals of diagnostic pathology*, 6(5), 281–287.
 40. Liao, X., Xin, X., & Lü, X. (2004). Primary Ewing's sarcoma-primitive neuroectodermal tumor of the vagina. *Gynecologic oncology*, 92(2), 684–688.
 41. McCluggage, W. G., Sumathi, V. P., Nucci, M. R., Hirsch, M., Dal Cin, P., Wells, M., Flanagan, A. M., & Fisher, C. (2007). Ewing family of tumours involving the vulva and vagina: report of a series of four cases. *Journal of clinical pathology*, 60(6), 674–680.

42. X. Ling, J. Tong, L. Wang, C. Yao, Z. Chen. (2021). Primary pulmonary Ewing's sarcoma: rare cause of massive hemothorax in a young girl-case report. *BMC Pediatr*, 21(1), 194.
43. Huh, J., Kim, K. W., Park, S. J., Kim, H. J., Lee, J. S., Ha, H. K., Tirumani, S. H., & Ramaiya, N. H. (2015). Imaging Features of Primary Tumors and Metastatic Patterns of the Extraskelatal Ewing Sarcoma Family of Tumors in Adults: A 17-Year Experience at a Single Institution. *Korean journal of radiology*, 16(4), 783–790.
44. Biermann J. S. (2013). Updates in the treatment of bone cancer. *Journal of the National Comprehensive Cancer Network : JNCCN*, 11(5 Suppl), 681–683.
45. Casali, P. G., Bielack, S., Abecassis, N., Aro, H. T., Bauer, S., Biagini, R., Bonvalot, S., Boukovinas, I., Bovee, J. V. M. G., Brennan, B., Brodowicz, T., Broto, J. M., Brugières, L., Buonadonna, A., De Álava, E., Dei Tos, A. P., Del Muro, X. G., Dileo, P., Dhooge, C., Eriksson, M., ... ESMO Guidelines Committee, PaedCan and ERN EURACAN (2018). Bone sarcomas: ESMO-PaedCan-EURACAN Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Annals of oncology : official journal of the European Society for Medical Oncology*, 29(Suppl 4), iv79–iv95.
46. Takahashi, D., Nagayama, J., Nagatoshi, Y., Inagaki, J., Nishiyama, K., Yokoyama, R., Moriyasu, Y., Okada, K., & Okamura, J. (2007). Primary Ewing's sarcoma family tumors of the lung a case report and review of the literature. *Japanese journal of clinical oncology*, 37(11), 874–877.
47. Siddiqui, M. A., Akhtar, J., Shameem, M., Baneen, U., Zaheer, S., & Shahid, M. (2011). Giant extraosseous Ewing sarcoma of the lung in a young adolescent female--a case report. *Acta orthopaedica Belgica*, 77(2), 270–273.
48. Ahmad R, Mayol BR, Davis M and Rougraff BT (1999): Extraskelatal Ewing's sarcoma. *Cancer* 85: 725 731.
49. Gaspar, N., Hawkins, D. S., Dirksen, U., Lewis, I. J., Ferrari, S., Le Deley, M. C., Kovar, H., Grimer, R., Whelan, J., Claude, L., Delattre, O., Paulussen, M., Picci, P., Sundby Hall, K., van den Berg, H., Ladenstein, R., Michon, J., Hjorth, L., Judson, I., Luksch, R., ... Oberlin, O. (2015). Ewing Sarcoma: Current Management and Future Approaches Through Collaboration. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*, 33(27), 3036–3046.
50. Demir, A., Gunluoglu, M. Z., Dagoglu, N., Turna, A., Dizdar, Y., Kaynak, K., Dilege, S., Mandel, N. M., Yilmazbayhan, D., Dincer, S. I., & Gurses, A. (2009). Surgical treatment and prognosis of primitive neuroectodermal tumors of the thorax. *Journal of thoracic oncology : official publication of the International Association for the Study of Lung Cancer*, 4(2), 185–192.
51. Shamberger, R. C., LaQuaglia, M. P., Gebhardt, M. C., Neff, J. R., Tarbell, N. J., Marcus, K. C., Sailer, S. L., Womer, R. B., Miser, J. S., Dickman, P. S., Perlman, E. J., Devidas, M., Linda, S. B., Krailo, M. D., Grier, H. E., & Granowetter, L. (2003). Ewing sarcoma/primitive neuroectodermal tumor of the chest wall: impact of initial versus delayed resection on tumor margins, survival, and use of radiation therapy. *Annals of surgery*, 238(4), 563–568.

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